

REVIEW ARTICLE

Brown fat and obesity: the next big thing?

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Summary

Brown adipose tissue (BAT) is well recognised to have an important role in the maintenance of body temperature in animals and human neonates, its thermogenic action affected by a tissue-specific uncoupling protein; fatty acid oxidation within the numerous brown adipocyte mitochondria is rendered inefficient leading to heat, rather than adenosine triphosphate (ATP), production. BAT was believed to show rapid involution in early childhood, leaving only vestigial amounts in adults. However, recent evidence suggests that its expression in adults is far more common than previously appreciated, with a higher likelihood of detection in women and leaner individuals. It is conceivable that BAT activity might reduce the risk of developing obesity since fat stores are used for thermogenesis, and a directed enhancement of adipocyte metabolism might have value in weight reduction. However, it is as yet unclear how such manipulation of BAT might be achieved; even in animal models, the control of thermogenic activity is incompletely understood. Even so, there is still much to interest the endocrinologist in BAT, with a range of hormones affecting adipocyte activity. This may either contribute to normal physiological function, or the phenotypical presentation of states of pathological hormone excess or deficiency. Thus, the gender differences in BAT distribution may be attributable to the differential effects of male and female sex hormones, whilst BAT expansion may drive the weight loss associated with catecholamine-producing pheochromocytomas. These observations support an important influence of the endocrine system on BAT activity and offer new potential targets in the treatment of obesity.

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Introduction

Obesity is a major global public health challenge. The prevalence of obesity has reached epidemic proportions and is expected to rise

further, such that the latest projections estimate that 2.16 billion overweight and 1.12 billion obese individuals will be affected globally by 2030. In addition to increased social stigmatization, financial disadvantage and impaired quality of life, obese individuals are faced with a markedly increased risk of cardiovascular disease, type 2 diabetes, osteoarthritis and a number of cancers. Lifestyle modification in the obese often provides only transient success, whilst clinical intervention has proven difficult; bariatric surgery is effective, but not without risk, and pharmaceutical options limited, especially following the recent withdrawal of both rimonabant and sibutramine within the European Union. The need for novel therapeutic approaches is therefore pressing. Brown adipose tissue (BAT), in contrast to white adipose tissue (WAT), is involved in energy dissipation rather than storage,¹ but was previously considered to have little physiological relevance in humans beyond early childhood. However, recent studies using ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) positron emission tomography–computed tomography (PET–CT) prove that BAT is present in adults, with activity notably declining with increasing obesity.² Working within the context of clinical endocrinology, the authors ask could the recruitment of larger amounts of BAT or an enhancement of its activity be an ‘antidote’ to obesity?

Energy balance and thermogenesis

Obesity is the product of a mismatch in energy supply and utilization, resulting in the deposition of WAT. In humans, this is almost inevitably because of a combination of excessive dietary intake and too little exercise. Interestingly, excision or denervation of BAT in animal models leads to an abnormal increase in WAT,¹ implying a significant impact upon energy balance. The primary function of BAT is in the generation of heat. Thermogenic mechanisms are customarily classified as either obligatory or facultative. Obligatory thermogenesis (OT) represents the energy dissipated as heat in the many energetic transformations inherent in life and equates to basal metabolic rate at thermoneutrality. In a cold environment, facultative thermogenesis (FT) may be required to maintain core temperature. Initially, heat is produced by shivering, which is replaced as acclimatization proceeds with nonshivering mechanisms in which BAT activation plays a key role. In small mammals, cold-induced thermogenesis in the absence of shivering accounts for an average of 11.8% of the resting metabolic rate, with high individual variation.³ Some observations have indicated that food intake results in a similar activation of brown adipocytes,⁴ which

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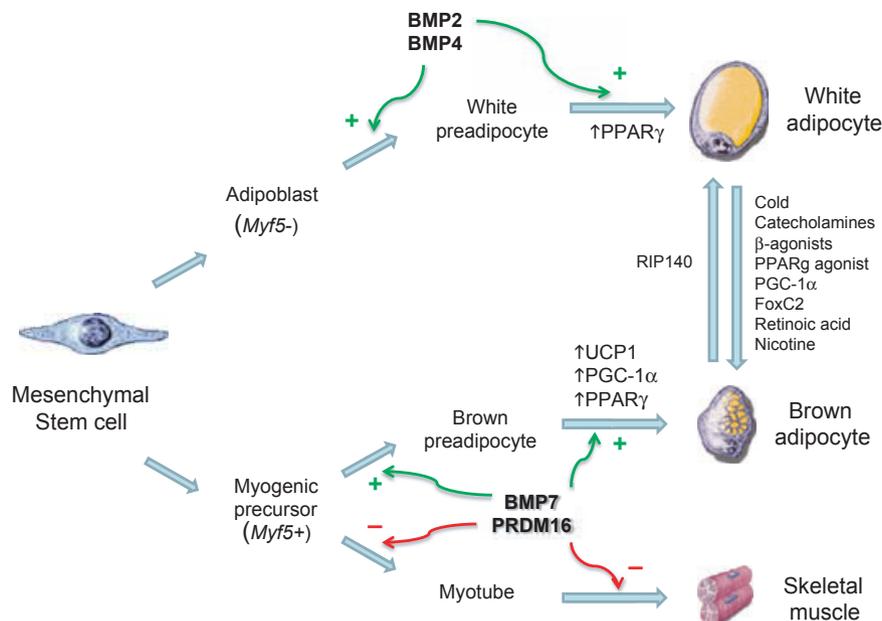


Fig. 1 Schematic representation of proposed developmental pathways of brown and white fat cells differentiation (where green arrows represent stimulatory influences, red arrows inhibitory). Bone morphogenic proteins 2 (BMP2) and 4 (BMP4) stimulate white adipocyte differentiation. BMP7 and PR-domain-containing 16 (PRDM16) stimulate brown adipocyte differentiation, whilst inhibiting promyogenic effectors, blocking potential development into skeletal muscle cells.

Table 1. Comparison of brown (BAT) and white (WAT) adipocyte morphology (modified from Fruhbeck *et al.*⁷³)

	BAT	WAT
Cell shape	Polygonal	Variable, but classically spherical
Cell size	Comparatively small (15–60 μm)	Variable, but large (25–200 μm)
Nucleus	Central, round or oval in shape	Peripheral, flattened
Cytoplasm	Large volume evenly distributed throughout cell	Thin, peripheral rim
Lipid content	Multiple small lipid droplets	Single large droplet occupying up to 90% of cell volume
Mitochondria	Abundant	Few
Endoplasmic reticulum (ER)	Present, but poorly developed	Little, but recognizable as rough and smooth ER
Tissue organization	Lobular, gland-like	Small lobules of densely packed cells
Cell content	Few other cell types	Multiple other cell types present
Vascularity	Highly vascularized	Adequate

has been taken as evidence in support of diet-induced thermogenesis. This envisages BAT acting not only to maintain body temperature but as a protector against obesity in times of positive energy balance, burning off excess calories in a co-ordinated mechanism

to maintain energy homeostasis. It is difficult to envisage the evolutionary drive for such a mechanism, whilst the most recent review expresses grave doubts about the existence of any such system.⁵

BAT structure and function

The ultimate phenotype of any cell is determined by the sequential activation of a cascade of transcription factors during differentiation. Those which drive the formation of WAT and BAT must inevitably activate expression of peroxisome proliferator-activated receptor- γ (PPAR γ), indispensable for adipogenesis. Whilst both WAT and BAT are derived from mesenchymal stem cells, perhaps unsurprisingly, they appear to have distinct lineages, with *Myf5* (shared with skeletal myocyte progenitors), *PGC-1 α* ⁶ and *PRDM16*⁷ (PR-domain-containing 16) expression, distinguishing the brown from white adipocyte precursors (see Fig. 1).

BAT and WAT adipocytes differ widely in morphology (summarized in Table 1), reflecting their different functional roles. Mitochondria are present in high numbers in brown adipocytes, particularly in comparison with white adipocytes, and are central to BAT activity. The mitochondria release chemical energy in the form of heat by means of uncoupling of oxidative phosphorylation, making the process of respiration inefficient. It is not intended to review the process in detail here, but in brief this phenomenon is mediated by uncoupling protein 1 (UCP1), which renders the inner membrane of the mitochondria ‘leaky’, and hence releases energy in the form of heat rather than storing it as ATP. UCP1 is in turn regulated by triiodothyronine (T3), possibly through the β -subunit of the thyroid hormone receptor, which is generated within BAT by the action of type 2 deiodinase (D2) on thyroxine (T4),

effectively creating a local, tissue-specific hyperthyroid state in the absence of changes in circulating thyroid hormones. Sympathetic stimulation, via the β_3 -adrenergic receptor (found almost exclusively in adipose tissue in rodents), enhances thermogenesis; the precise mechanism by which this is achieved remains unclear, however.

Whilst both BAT and WAT, as would be expected, have neuro-anatomically well-characterized sympathetic innervation (with activation initiating thermogenesis and lipid mobilization, respectively), there is little evidence to support the presence of a putatively counter-regulatory parasympathetic input.⁸ Previous studies have however hinted indirectly at such, atropine, a nonspecific muscarinic receptor (MR) antagonist, has been shown to enhance the thermogenic response in rats,⁹ although the authors failed to demonstrate the presence of any markers of parasympathetic innervation at that time. Interestingly, despite a lack of evidence to support this counter-regulatory role for the parasympathetic nervous system, M3 MR knockout mice weigh significantly less (22%) than wild-type, attributable to a marked reduction in visceral WAT;¹⁰ no indication of BAT status was provided.

In most mammals, BAT is found predominantly in anatomically discrete depots, classically the interscapular region and axillae (for review, see Cannon & Nedergaard¹¹). However, BAT can emerge in animal WAT depots in response to cold or prolonged β -adrenergic stimulation, but the mechanism remains unclear. Certainly, brown adipocytes have been identified histologically in up to 50% of younger patients, seeded amongst white adipocytes,¹² whilst based on UCP1 mRNA quantification,¹³ there would appear to be approximately one brown adipocyte per 100–200 white adipocytes within human visceral WAT. Thus, the observed increase in BAT may be because of simple clonal expansion and maturation of an already present population of committed preadipocytes. However, it has been recently reported that rat epididymal WAT adipocytes chronically exposed to the thiazolidinedione (TZD) rosiglitazone (a PPAR γ agonist) share BAT-type characteristics, but do not express the transcription factors (most notably PRDM16) associated with BAT.¹⁴ These appear to be the so-called *brite* (brown-in-white) cells, which have BAT characteristics, but do not seem to have the same lineage. This report suggests that these brite cells arise from trans-differentiation. The possibility of a pharmacologically driven switch from WAT to BAT is highly attractive in the context of the clinical management of obesity.

Is there functional BAT in humans?

Or, perhaps more specifically, is there functional BAT in *adults*? Whilst present in significant quantities in the neonate,¹⁵ until recently, it was assumed that rapid involution in the first years of life left only vestigial amounts of BAT in healthy human adults. Physiologically, the requirement for nonshivering thermogenesis is limited by our other adaptations to environment (clothing and heating), although a study nearly 30 years ago showed at least one group of outdoor workers in Scandinavia to have increased BAT deposits.¹⁶ Whilst information about normal tissue function can often be gleaned from pathological states, this also proved to be of limited value for BAT. Brown fat tumours (or *hibernomas*, from



Fig. 2 ^{18}F -FDG-PET image demonstrating brown adipose tissue deposits in the supraclavicular, cervical and mediastinal sites.

their resemblance to adipose tissue in hibernating animals¹⁷) are rare, benign and seemingly asymptomatic; though, significant weight loss has been associated with the tumour in at least one case report.¹⁸ Interestingly, β_3 -adrenoreceptor polymorphisms leading to a reduction in receptor function (notably the substitution of a tryptophan to an arginine residue at position 64) have been linked to weight gain,¹⁹ as well as being positively associated with early-onset type 2 diabetes.²⁰

Circumstantial evidence for BAT activity came from the experience of those interpreting PET–CT scans (see Fig. 2). The radio-tracer ^{18}F -FDG follows glucose metabolism initially, but does not enter the Krebs cycle after phosphorylation, and is therefore trapped within the cell. ^{18}F -FDG-PET simply monitors glucose uptake, with highly metabolically active tissues being labelled; what this actually means can, of course, only be inferred. It was noted that there was confounding high symmetrical uptake in regions previously found consistent with postmortem locations of BAT and that the intensity of these signals could be reduced by increasing the ambient temperature or by β -blockade. In the first half of 2009, five independent groups used ^{18}F -FDG-PET to identify and characterize BAT in adult humans (see Table 2 for summary). In a retrospective analysis of 3640 consecutive scans from 1972 patients, Cypess *et al.*² found putative BAT deposits in 7.5% of women and 3.1% of men, located mainly in the cervical, supraclavicular, axillary and paravertebral regions (echoing the sites in the neonate). As well as being present in more than twice as many women, the deposits were also larger and showed greater levels of ^{18}F -FDG uptake in female compared to male subjects. Probability of

Table 2. Summary of recent reports supporting the presence of functional BAT in humans

Authors	Methods	Results
Cypess <i>et al.</i> ²	¹⁸ F-FDG-PET	Retrospective 3640 consecutive scans (1972 patients) 106 (5.4%) with BAT Higher prevalence in women Lower prevalence with increasing age Seasonal variation β -blockers reduced probability of detection
Van Marken Lichtenbelt <i>et al.</i> ²¹	¹⁸ F-FDG-PET	24 young (18–32 years) males Two groups (normal and high BMI) 23 (96%) with BAT Cold-induced increased ¹⁸ F-FDG uptake Lower uptake with increasing BMI
Virtanen <i>et al.</i> ²²	¹⁸ F-FDG-PET biopsy	5 male volunteers Cold-induced increased ¹⁸ F-FDG uptake Paired biopsies (putative BAT; WAT control) UCP1 (mRNA/protein) in BAT only PRDM16, PPAR γ , β 3AR, D2 (mRNA) in higher concentrations than WAT
Zingaretti <i>et al.</i> ²³	Biopsy	8 male and 27 females Neck fat biopsied during thyroid surgery 10 (28%) positive for UCP1 protein UCP1 positivity associated with lower BMI
Saito <i>et al.</i> ²⁴	¹⁸ F-FDG-PET	31 male and 25 female volunteers Age range 23–65 years Cold-induced increased ¹⁸ F-FDG uptake Lower uptake with increasing BMI Seasonal variation

BAT, brown adipose tissue; BMI, body mass index; PET, positron emission tomography; ¹⁸F-FDG, ¹⁸F-fluorodeoxyglucose; PPAR γ , proliferator-activated receptor- γ ; WAT, white adipose tissue.

detection was inversely associated with age of subject, body mass index (BMI), β -blocker use and outdoor temperature at the time of the scan. Confirming the importance of temperature on BAT activity, Van Marken Lichtenbelt *et al.*²¹ demonstrated increased metabolic activity in (putative) BAT predominantly in the neck and supraclavicular region of 23 of 24 young, healthy male subjects

when exposed to mild cooling. Activity showed high intersubject variability. Mean activity was significantly lower in overweight or obese (BMI > 25) subjects. Interestingly, the single subject in whom there was no demonstrable ¹⁸F-FDG uptake also had the highest BMI (38.7). Virtanen *et al.*²² reported a 15-fold increase in ¹⁸F-FDG uptake under similar conditions, but were also able to confirm the presence of BAT in these regions by demonstrating the presence of UCP1 mRNA and protein in biopsies from three of their five subjects. In addition, they showed the expression of D2 and the β 3-adrenergic receptor, further indicating the potential for function. This potential was supported by the demonstration of distinct islands of BAT in the necks of approximately one-third of a group of 35 patients undergoing thyroid surgery by Zingaretti *et al.*²³ The islands were highly vascular, with a rich sympathetic innervation, and demonstrated the presence of brown adipocyte precursors. Whilst also confirming the inverse relationship between suprascapular and paraspinal ¹⁸F-FDG uptake and both total and visceral adiposity in their group of 56 healthy volunteers in response to acute cold, Saito and co-workers²⁴ demonstrated a degree of seasonal variability to their findings, with increased uptake during the winter months.

It would be reasonable to conclude that cold-inducible BAT is present in adults, with the largest depots in and around the neck. Observed in leaner individuals, and inversely associated with other indices of the metabolic syndrome, it may be implied that increasing BAT mass or activity offers a degree of protection from obesity. This is particularly so in the light of the relatively limited amounts of such tissue that would be required to make significant impact on energy balance; it has been estimated that as little as 50 g of BAT would account for 20% of daily energy expenditure.²⁵ Based upon the registered rate of ¹⁸F-FDG uptake and data from substrate preference in rodents,²⁶ Virtanen and co-workers²² speculated that the estimated 63 g of BAT found in the supraclavicular/paracervical depot of one of the subjects could combust the energy equivalent of 4.1 kg of WAT over 1 year. Before consideration of the therapeutic possibilities of this phenomenon, it may be of value to consider BAT in the context of wider endocrine function and disease in both animal models and humans.

Clinical aspects of hormonal modulation of BAT activity: points for endocrinologists

BAT is a potential target for a wide variety of hormones physiologically. It is thus possible that BAT activity may be affected where hormone levels are pathologically elevated or suppressed. Starting with the catecholamines and thyroid hormones, which, as noted above, are vital to BAT activity, this section will briefly examine the direct effects of the clinically more important hormones on BAT, utilizing data from human and animal studies with particular reference to endocrine disease.

BAT and pheochromocytomas

In the light of the key role of adrenergic stimulation in thermogenesis, one might predict significant expansion of BAT in patients with catecholamine-producing pheochromocytomas. An associa-

tion with brown fat tumours was noted nearly 40 years ago,²⁷ whilst Lean and co-workers¹⁵ demonstrated the presence of BAT histologically, not present in controls, following laparotomy in three patients with pheochromocytomas.

Enhanced ¹⁸F-FDG uptake, which disappeared following surgical resection, has been reported in a patient with an extra-adrenal pheochromocytoma,²⁸ emphasizing the effect of excess circulating catecholamines in enhancing BAT activity; interestingly, Hadi *et al.*²⁹ have demonstrated that the likelihood of identifying BAT in this patient group on PET-CT is increased at higher plasma noradrenaline levels.

BAT and thyroid status

A recent case report from Skarulis *et al.*³⁰ appears to link BAT activity and volume with circulating thyroid hormone levels. Classically of course, thyroid over- or underactivity gives rise to distinct phenotypes, reflecting the contrasting effects of hormone excess or deficiency on metabolism, body mass and composition. Thus, thyrotoxic individuals are heat intolerant and lean with reduced body fat, whilst hypothyroidism is associated with sensitivity to low ambient temperatures and weight gain. Hypothyroidism because of thyroid-stimulating hormone resistance is associated with an inability to maintain core temperature when exposed to cold in a mouse model (*hyt/hyt*) lacking a functional thyroid-stimulating hormone receptor; this can be reversed by gene transfer into BAT.³¹ Conversely, as with pheochromocytomas, some of the clinical features of hyperthyroidism could conceivably be explained in terms of an increased activation of BAT-dependent FT, but in this instance owing to excess circulating T3. Previously, it had been argued that the requirement for a concomitant β 3-adrenergic activation was against this; sympathetic stimulation of BAT had been described as being inversely related to thyroid status and thus reduced in the presence of excess systemic thyroid hormones (reviewed in Silva³²). In addition, high concentrations of T4 themselves were shown to powerfully inhibit D2 activity.³³ Thus, it was felt that a generalized enhancement of metabolism, and thus increase in OT such as seen in skeletal muscle,³⁴ was the basis of the development of the hyperthyroid phenotype, rather than an upregulation of BAT activity.

However, Lopez *et al.*³⁵ (2010) have recently suggested that elevated T3 levels do indeed activate BAT, but indirectly via action at the ventromedial nucleus of the hypothalamus (VMH). They demonstrated that high levels of circulating thyroid hormones, or central administration of T3, were associated with enhanced neuronal activation and β 3-adrenergic signalling, a shift in the ratio of BAT to WAT within fat pads, and increased BAT UCP1, PPAR γ and D2 expression in rats. The key process appears to be a reduction in the activity of hypothalamic adenosine monophosphate-activated protein kinase. This leads to changes in lipid metabolism within the VMH, although it is not yet clear how this and the observed increased activity of nerves innervating BAT are related. It should be noted that no direct measure of BAT activity was used (enhancement implied by increased expression of thermogenic markers) nor any assessment of metabolic rate made (which would help to address the impact of OT in this context). Though one should

always be cautious when extrapolating from animal models, a similar mechanism may operate in humans, and would certainly be compatible with the observations reported by Skarulis and co-workers.³⁰ This case appears somewhat atypical however, exhibiting significant confounding features. BAT changes were observed in a clinically euthyroid patient receiving supraphysiological doses of thyroxine to suppress thyroid-stimulating hormone (TSH) levels on a background of papillary thyroid carcinoma and insulin resistance secondary to insulin receptor mutation. Although there was an overall reduction in weight and an improvement in glycaemic control with thyroxine treatment, there was no change in relative fat mass, whilst the contribution of obligate thermogenic mechanisms was only partially addressed. There is no evidence to support the therapeutic use of T3 to stimulate thermogenesis in obese individuals.

Whether BAT activation is or is not directly responsible for some of the signs and symptoms of thyroid disease, one still might predict there to be an association between the relative levels of circulating thyroid hormones (which exhibit a relatively wide 'normal range' in clinically euthyroid individuals) and degree of obesity (as a surrogate marker of BAT volume). As well as its autocrine effect on UCP1 production, there is a net release of T3 from BAT into the circulation.³⁶ This can be of considerable physiological significance; in rodents, for example, it has been suggested that BAT may be responsible for about half of the total systemic conversion of T4 to T3.³⁷ BAT deposits in humans might be expected to behave similarly. Intriguingly, Shon *et al.*³⁸ have recently reported a negative correlation between plasma-free T4 levels and BMI in a large (1572) female cohort. Unfortunately, only T4 is discussed, although De Pergoli *et al.*³⁹ had previously shown free T3 levels to be positively associated with both BMI and waist circumference in a similar, though, smaller (201 women) experimental group. In addition, they reported TSH levels to also be positively associated with waist circumference, in agreement with the observations of Iacobellis *et al.*⁴⁰ Shon and co-workers³⁸ failed to show any such associations for TSH. These observations may be interpreted as showing an enhanced T3 generation leading to relative T4 depletion and thus increased TSH signalling. As this occurs in the context of increasing obesity, enhanced BAT D2 activation is an unlikely source of this excess T3 production. Interestingly, bolus leptin has been reported to increase type 1 deiodinase (D1) activity in both the thyroid and liver of euthyroid rats;⁴¹ it may be speculated that the chronically increased plasma concentrations observed in obese individuals could have a comparable effect and thus underlie the observed variation in thyroid hormone levels.

BAT and corticosteroids

Clinically, corticosteroid excess (Cushing's syndrome) is associated with obesity, primarily owing to an increase in visceral adipose tissue.⁴² In animal models, this is driven by an expansion of WAT, which appears to be more sensitive to corticosteroids than BAT.⁴³ Both WAT and BAT express functional glucocorticoid receptors.⁴⁴ Interestingly, the reported effects of corticosteroids on BAT would be consistent with a reciprocal down-regulation of brown adipocyte activity. Thus, noradrenaline-induced UCP1 mRNA

accumulation in a BAT cell line has been shown to be reduced by both an endogenous and synthetic corticosteroid,⁴⁵ whilst excess steroid increases the number and size of lipid droplets in brown adipocytes in a dose-dependent manner.⁴³ An inhibitory effect on β 3-adrenoreceptor expression *in vitro* has also been reported, although this appears to be only transient *in vivo*, despite continued exposure to corticosteroids.⁴⁶ Ashizawa and co-workers⁴⁷ demonstrated that the plasma levels of both leptin and adiponectin changed predictably as obesity improved following adrenalectomy in a patient with Cushing's syndrome. This may simply correlate to the observed loss of mass from WAT deposits, rather than any direct causal relationship. Conversely, Addison's disease (corticosteroid deficiency) may have only a limited effect on BAT; Berthiaume and co-workers reported adrenalectomy to have no effect on BAT mass in rats, despite a marked reduction in visceral WAT.⁴⁸

BAT and the sex hormones

The sex hormones have a marked effect on BAT activity. Receptor expression differs between BAT of male and female origin, with higher numbers of all receptor types in male brown adipocytes.⁴⁹ Rodriguez *et al.*⁵⁰ reported that cultured brown adipocytes, when exposed to testosterone, had fewer, smaller lipid droplets than untreated cells and expressed lower levels of UCP1 mRNA in response to adrenergic stimulation; a concomitant reduction in PGC1 α transcription has also been described.⁵¹ The precise mechanism by which testosterone down-regulates BAT activity remains unclear, although it induces increased expression of the antilipolytic α 2A-adrenoreceptor in brown adipocytes.⁵²

As perhaps might be predicted, the female sex hormones appear to enhance BAT function. Certainly, oestrogen deficiency has long been noted to reduce thermogenic capacity in rats following ovariectomy,⁵³ associated with a reduction in BAT UCP1 levels.⁵⁴ The reported effects of the oestrogens and progesterone on brown adipocytes often overlap, but not invariably so; thus, whilst both promote the formation of larger, more numerous lipid droplets, only progesterone is noted to increase UCP1 mRNA levels.⁵⁰ The female hormones (predominantly progesterone) also influence adrenergic stimulation of BAT by modulating the numbers of cell surface receptors, down-regulating (in contrast to testosterone) α 2A- and up-regulating β 3-adrenoreceptors⁵² (although the affinity of the latter for noradrenaline may simultaneously be reduced⁵⁵). In an additional boost to BAT activity, the female sex hormones also appear to positively influence mitochondrial production and recruitment.⁵¹

It is possible that it is not only the sex hormones themselves that effect BAT activity. Dehydroepiandrosterone (DHEA), a sex hormone precursor usually present in plasma in its sulphated form, is directly able to inhibit proliferation of a human brown adipocyte cell line,⁵⁶ perhaps as a necessary preliminary to differentiation. Supplementation in obese rats (which are relatively deficient in DHEAS) increases PGC1 α , UCP1 and β 3-adrenoreceptor expression; though, this was not seen in similarly treated lean rats.⁵⁷ The different effects of the male and female sex hormones on adipose tissue may thus have a significant impact on phenotype, whether this be the gender-specific distribution of WAT (predominantly

visceral in males and subcutaneous in females⁵⁸) or the increased incidence of functional BAT in females described in the recent ¹⁸F-FDG-PET studies.

BAT and the GH/IGF-1 axis

The elevated plasma growth hormone (GH) and IGF-1 levels of active acromegaly are clinically associated with increased lean body mass and reduced fat,⁵⁹ whilst treatment of athletes with GH has similar effects on body composition.⁶⁰ Indeed these effects have been utilized therapeutically in the treatment of HIV-associated adipose redistribution syndrome, a disorder characterized by excess truncal WAT.⁶¹ It is possible that, in addition to this mobilization of WAT stores, there is a complementary increase in BAT activity, although this has not been clearly demonstrated in patients with acromegaly. Certainly, transgenic mouse models in which GH function has been disrupted or augmented have expanded or reduced interscapular BAT depots, respectively.⁶² However, whilst treatment of obese mice with exogenous GH reduced both visceral and subcutaneous WAT in a dose-dependent fashion, there was no expansion of the interscapular BAT;⁶³ though, at higher doses of GH, a modest (twofold) increase in BAT UCP1 mRNA was noted. Interestingly, a sixfold increase in WAT UCP1 mRNA was also observed. This may represent brite cell induction; though, it may simply be that the small, pre-existing population of brown adipocytes become relatively more prominent as adiposity decreases; certainly, treated lean mice, which show no significant change in body composition, do not demonstrate any UCP1 increase.

IGF-1 receptors are present in high numbers on the plasma membrane of brown preadipocytes; stimulation leads to mitogenesis in cultured foetal mouse BAT cells⁶⁴ and protects against apoptosis.⁶⁵ Cold exposure induces increased IGF-1 mRNA expression in rats,⁶⁶ suggesting the possibility of autocrine stimulation in response to a fall in temperature. This appears to be transient however,⁶⁷ peaking in the first few days after the onset of cold, and thus likely to be of significance only in the initial phase of acclimatization. Interestingly, IGF-1 also upregulates type III iodothyronine 5-deiodinase expression in BAT adipocytes,⁶⁸ which would enhance T3 inactivation with predictable effects on the activity and thermogenic capability of the cell. Thus, IGF-1 action is potentially self-limiting, whether in response to cold or excess plasma levels. The direct effects of GH on BAT are equivocal, with little discernible increase in activity even in excess. It is therefore unlikely that BAT is a target of the GH/IGF-1 axis.

BAT and the endocannabinoids

The clinical significance of the endocannabinoid system stems essentially from the effects of rimonabant on weight gain in diabetics. As well as regulating energy balance by modulating hypothalamic circuits which control food intake and energy expenditure, there is now considerable evidence that the endocannabinoids act directly on adipose tissue (reviewed in Vettor & Pagano⁶⁹). There is functional expression of cannabinoid receptor type 1 (CB1) in both rodent and human white adipocytes (although expression of CB2 remains controversial); activation enhances glucose uptake and fat

droplet accumulation. Rimonabant is a CB1 receptor antagonist; chronic usage increases interscapular BAT temperature and upregulates UCP1.⁷⁰ Both effects are attenuated by denervation of the tissue, suggesting the effect is mediated, at least in part, by increased sympathetic activity. However, a recent report of rimonabant also inducing UCP1 production and upregulating mitochondrial numbers in cultured WAT adipocytes⁷¹ implies an end-organ action. Similarly, the increase in serum adiponectin levels associated with rimonabant treatment is only partially accounted for by weight loss, further suggesting a direct effect on adipocytes. Whilst there is, as yet, no report of BAT CB1 expression, this would seem likely, with activation inhibiting BAT activity, whilst promoting WAT activity.

Is therapeutic manipulation of BAT possible?

Although championed on occasion as having potential as an anti-obesity organ,⁷² the assumption that BAT was present only in neonates limited the appeal of any such approach. However, the recent demonstrations of significant deposits of adult BAT reviewed earlier have once again raised the possibility of their therapeutic manipulation to promote weight reduction (for review, see Fruhbeck *et al.*⁷³). Although increased energy expenditure does not necessarily guarantee weight loss (just as a low basal metabolic rate does not necessarily lead to obesity⁷⁴), it is precisely this, via an enhancement of FT, which is the ultimate goal of any BAT-orientated strategy to combat obesity in humans. There are a series of complementary facets that might contribute to any successful intervention; thus, one might aim to stimulate the activity of already existing BAT, whilst favouring the recruitment of the few brown adipocytes in WAT or promoting the emergence of new BAT depots.

As might be expected, this is not entirely straightforward using traditional pharmacological approaches, owing to the lack of suitable pharmaceutical agents, and problems with unwanted additional effects. Take, for example, the use of adrenergic agents. Whilst β 3-adrenergic stimulation is required to activate thermogenesis, proliferatively competent brown adipocytes express β 1- rather than β 3-receptors. Exogenous β 1 stimulation is thus required to increase cell numbers; this, unfortunately, brings with it unacceptable consequences on cardiac function. Similarly, the promising action of the TZD compounds has to be balanced against the possibly increased risk of cardiac and bone side effects. Although this remains controversial, the use of rosiglitazone is already proscribed. Rimonabant has similarly been withdrawn in Europe owing to an increased risk of serious psychiatric side effects. Retinoic acid induces UCP1 expression in both BAT and WAT,⁷⁵ but has a wide range of other effects *in vivo*, limiting utilization.

If the use of pharmaceutical agents is currently challenging, the induction of specific gene expression to enhance BAT levels and activity has some merit. It has been demonstrated that enhancement of UCP1 levels by direct overexpression produces mice that are resistant to genetic- or diet-induced obesity (reviewed in Kozak & Koza⁷⁶); the loss of inhibitors of UCP uncoupling activity [such as *Cidea* (cell death-inducing DFF45-like effector A)]⁷⁷ produces similar results. The overexpression of *FOXC*₂ (a winged helix transcription factor) exclusively in adipocytes forces the shift from

WAT to BAT in transgenic mice⁷⁸ as β 3-adrenergic receptors are upregulated and cAMP levels increased. The inhibition of RIP140, a nuclear corepressor, shows similar promise. Known to repress UCP1 expression, RIP140-null adipocytes exhibit increased levels of UCP1 mRNA.⁷⁹

Initially, the authors asked whether recruitment of larger amounts of BAT or an enhancement of its activity could be an 'antidote' to obesity – perhaps, it is now more pertinent to ask whether this is actually practically possible. Whilst a pharmaceutical approach is clearly preferable economically, the lack of readily available candidates precludes this currently. In most cases, even where genetic approaches show promise, they tend to remain primarily research tools rather than progressing to therapies; the treatment of obesity is potentially so lucrative that it may prove to be an exception.

Conclusions

As the problems presented by obesity and its associated morbidities continue to grow, it appears that, paradoxically, adipose tissue itself may provide a solution. The adipose organ is not homogeneous, but comprises of two major tissue types of distinctive structure, function and lineage, as evidenced by their differing gene expression profiles; thus, BAT resembles muscle, but WAT is akin to macrophages.^{80,81} White adipose tissue performs the task most readily associated with adipose tissue, that of energy storage (though in a less passive fashion than had, perhaps, been believed previously), whilst the function of BAT is to protect body temperature in response to environmental cooling, generating heat by inefficient mitochondrial metabolism. Whilst these two types are well preserved in most animals, BAT had been thought to be restricted to neonates only in man. Whilst, as a benefit of progress, man has acquired other means to keep himself warm, this has come with a relative abundance of food, a consistently positive energy balance, and thus endemic obesity. Obesity management is fraught with difficulties ranging from the limited range and inadequacy of therapeutic options to poor patient adherence with lifestyle changes. The recent demonstrations of significant deposits of functional adult human BAT suggest another approach, namely the manipulation of facultative thermogenesis as an aid to weight loss. Whilst increasing both brown adipose tissue volume and activity is theoretically feasible (and is seen pathologically, on occasion), there are still many practical issues that require attention. It is however safe to say that we live in exciting times, particularly if that last diet just did not work!

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Addendum

The articles in this review were generated by multiple searches of the PubMed and Medline databases utilizing the keywords 'brown

adipose tissue' either alone or in conjunction with the specified hormones or pathological states.

References

- Hansen, J.B. & Kristiansen, K. (2006) Regulatory circuits controlling white versus brown adipocyte differentiation. *Biochemical Journal*, **398**, 153–168.
- Cypess, A.M., Lehman, S., Williams, G. *et al.* (2009) Identification and importance of brown adipose tissue in adult humans. *New England Journal of Medicine*, **360**, 1509–1517.
- Claessens-van Ooijen, A.M., Westerterp, K.R., Wouters, L. *et al.* (2006) Heat production and body production during cooling and rewarming in over-weight and lean men. *Obesity*, **14**, 1914–1920.
- Wijers, S.L.J., Sarjis, W.H.M. & van Marken Lichtenbelt, W.D. (2007) Individual thermogenic responses to mild cold and over-feeding are closely related. *Journal of Clinical Endocrinology and Metabolism*, **92**, 4299–4305.
- Kozak, L.P. (2010) Brown fat and the myth of diet-induced thermogenesis. *Cell Metabolism*, **11**, 263–267.
- Seale, P., Kajimura Yang, S.W. *et al.* (2007) Transcriptional control of brown fat determination by PRDM16. *Cell Metabolism*, **6**, 38–54.
- Uldry, M., Yang, W., St Pierre, J. *et al.* (2006) Complementary action of PGC1 coactivation in mitochondrial biogenesis and brown fat differentiation. *Cell Metabolism*, **3**, 333–341.
- Bartness, T.J. & Song, C.K. (2007) Sympathetic and sensory innervation of white adipose tissue. *Journal of Lipid Research*, **48**, 1655–1672.
- Bryant, K.R., Rothwell, N.J., Stock, M.J. *et al.* (1983) Parasympathetic effects on diet-induced thermogenesis. *European Journal of Pharmacology*, **95**, 291–294.
- Yamada, M., Miyakawa, T., Duttaroy, A. *et al.* (2001) Mice lacking the M3 muscarinic acetylcholine receptor are hypophagic and lean. *Nature*, **410**, 207–212.
- Cannon, B. & Nedergaard, J. (2004) Brown adipose tissue: function and physiological significance. *Physiological Reviews*, **84**, 277–359.
- Nedergaard, J., Bengtsson, T. & Cannon, B. (2007) Unexpected evidence for active brown adipose tissue in adult humans. *American Journal of Physiology. Endocrinology and Metabolism*, **293**, E444–E452.
- Oberkofler, H., Dallinger, G., Liu, Y.M. *et al.* (1997) Uncoupling protein gene: quantification of expression levels in adipose tissue of obese and non-obese humans. *Journal of Lipid Research*, **38**, 2125–2133.
- Petrovic, N., Walden, T.B., Shabalina, I.G. *et al.* (2010) Chronic PPAR γ activation of epididymally derived white adipocyte cultures reveals a population of thermogenically competent, UCP 1-containing adipocytes molecularly distinct from classical brown adipocytes. *The Journal of Biological Chemistry*, **285**, 7153–7164.
- Lean, M.E., James, W.P., Jennings, G. *et al.* (1986) Brown adipose tissue in patients with phaeochromocytoma. *International Journal of Obesity*, **10**, 219–227.
- Huttenen, P., Hirvonen, J. & Kinnula, U. (1981) The occurrence of brown adipose tissue in outdoor workers. *European Journal of Applied Physiology and Occupational Physiology*, **46**, 339–345.
- Furlong, M.A., Famburg-Smith, J.C. & Miettinen, M. (2001) The morphological spectrum of hibernomas in a clinico-pathological study of 170 cases. *American Journal of Surgical Pathology*, **25**, 809–814.
- Allegra, S.R., Gmuer, C. & O' Leary, G.P. (1983) Endocrine activity in a large hibernoma. *Human Pathology*, **14**, 1044–1052.
- Iacobellis, G., Ribaldo, M.C., Zappaterreno, A. *et al.* (2005) Relationship of thyroid function with body mass index, leptin insulin sensitivity and adiponectin in euthyroid obese women. *Clinical Endocrinology*, **62**, 487–491.
- Cabanelas, A., Lisboa, P.C., Moura, E.G. *et al.* (2007) Acute effects of leptin on 5'-deiodinases are modulated by thyroid state of fed rats. *Hormone and Metabolic Research*, **39**, 818–822.
- Van Lichtenbelt Marken, W.D., Vanhommerig, J.W., Smulders, N.M. *et al.* (2009) Cold-activated brown adipose tissue in healthy men. *New England Journal of Medicine*, **360**, 1500–1508.
- Virtanen, K.A., Lidell, M.E., Orava, J. *et al.* (2009) Functional brown adipose tissue in healthy adults. *New England Journal of Medicine*, **360**, 1518–1525.
- Zingaretti, M.C., Crosta, F., Vitali, A. *et al.* (2009) The presence of UCP-1 demonstrates that metabolically active adipose tissue in the neck of adult humans truly represents brown adipose tissue. *FASEB Journal*, **23**, 3113–3120.
- Saito, M., Okamatsu-Ogura, Y., Matsushita, M. *et al.* (2009) High incidence of metabolically active brown tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes*, **58**, 1526–1531.
- Rothwell, N.J. & Stock, M.J. (1983) Luxuskonsumtion, diet-induced thermogenesis and brown fat: the case in favour. *Clinical Science*, **64**, 19–23.
- Ma, S.W. & Foster, D.O. (1986) Uptake of glucose and release of fatty acids and glycerol by rat brown adipose tissue in vitro. *Canadian Journal of Physiology and Pharmacology*, **64**, 609–614.
- English, J.T., Patel, S.K. & Flanagan, M.J. (1973) Association of phaeochromocytomas with brown fat tumours. *Radiology*, **107**, 279–281.
- Ramacciotti, C., Schneegans, O., Lang, H. *et al.* (2006) Diffuse uptake of brown fat on computed-tomography coupled positron emission tomocinitgraphy (CT-PET) for the exploration of extra-adrenal phaeochromocytoma. [French]. *Annales d'endocrinologie*, **67**, 14–19.
- Hadi, M., Chen, C.C., Whatley, M. *et al.* (2007) Brown fat imaging with (18)F-6-fluorodopamine PET/CT, (18)F-FDG PET/CT, and (123)I-MIBG SPECT: a study of patients being evaluated for phaeochromocytoma. *Journal of Nuclear Medicine*, **48**, 1077–1083.
- Skarulis, M.C., Celi, F.S., Mueller, E. *et al.* (2010) Thyroid hormone induced brown adipose tissue and amelioration of diabetes in a patient with extreme insulin resistance. *Journal of Clinical Endocrinology and Metabolism*, **95**, 256–262.
- Endo, T. & Kobayashi, T. (2008) Thyroid stimulating hormone receptor in brown adipose tissue is involved in the regulation of thermogenesis. *American Journal of Physiology*, **295**, E514–E518.
- Silva, J. (2000) Catecholamines and the sympathoadrenal system in thyrotoxicosis. In: L.E. Braverman, R.D. Utiger eds. *Werner and Ingbar's The Thyroid: A Fundamental and Clinical Text*. Lippincott Williams & Wilkins, Baltimore, USA, 642–651.
- Silva, J.E. & Larsen, P.R. (1986) Interrelationships among thyroxine, growth hormone, and the sympathetic nervous system in the regulation of 5'-iodothyronine deiodinase in rat brown adipose tissue. *Journal of Clinical Investigation*, **77**, 1214–1223.
- Silva, J.E. (2006) Thermogenic mechanisms and their hormone regulation. *Physiological Reviews*, **86**, 435–464.

- 35 Lopez, M., Varela, L., Vasquez, M.J. *et al.* (2010) Hypothalamic AMPK and fatty acid metabolism mediate thyroid regulation of energy balance. *Nature Medicine*, **16**, 1001–1008.
- 36 Fernandez, J.A., Mampel, T., Villarroja, F. *et al.* (1987) Direct assessment of brown adipose tissue as a site of systemic triiodothyronine production in rat. *Biochemical Journal*, **243**, 281–284.
- 37 Silva, J.E. & Larsen, P.R. (1985) Potential of brown adipose tissue type II thyroxine 5'-deiodinase as a local and systemic source of triiodothyronine in rats. *Journal of Clinical Investigation*, **76**, 2296–2305.
- 38 Shon, H.S., Jung, E.D., Kim, S.M. *et al.* (2008) Free T4 is negatively correlated with body mass index in euthyroid women. *The Korean Journal of Internal Medicine*, **23**, 53–57.
- 39 De Pergoli, G., Ciampolillo, A., Paolotti, S. *et al.* (2007) Free triiodothyronine and thyroid stimulating hormone are directly associated with waist circumference, independently of insulin resistance, metabolic parameters and blood pressure in overweight and obese women. *Clinical Endocrinology*, **67**, 265–269.
- 40 Clement, K., Vaisse, C., Manning, B.S. *et al.* (1995) Genetic variation in the β 3-adrenergic receptor and an increased capacity to gain weight in patients with morbid obesity. *The New England Journal of Medicine*, **333**, 352–354.
- 41 Walston, J., Silver, K., Bogardus, C. *et al.* (1995) Time of onset of non-insulin dependent diabetes mellitus and genetic variation in the β 3-adrenergic receptor gene. *The New England Journal of Medicine*, **333**, 343–347.
- 42 Geer, C.B., Shen, W., Gallagher, D. *et al.* (2010) MRI assessment of lean and adipose tissue distribution in female patients with Cushing's disease. *Clinical Endocrinology*, **73**, 469–475.
- 43 Strack, A.M., Bradbury, M.J., Dallman, M.F. *et al.* (1995) Corticosterone decreases non-shivering thermogenesis and increases lipid storage in brown adipose tissue. *American Journal of Physiology*, **268**, R183–R191.
- 44 Feldman, D. & Loose, D. (1977) Glucocorticoid receptors in adipose tissue. *Endocrinology*, **100**, 398–405.
- 45 Soumano, K., Desbiens, S., Rabelo, R. *et al.* (2000) Glucocorticoids inhibit the transcriptional response of the uncoupling protein 1 gene to adrenergic stimulation in a brown adipose cell line. *Molecular and Cellular Endocrinology*, **165**, 7–15.
- 46 Bakopanos, E. & Silva, J.E. (2002) Opposing effects of glucocorticoids on β 3-adrenergic receptor expression in HIB-1B brown adipocytes. *Molecular and Cellular Endocrinology*, **190**, 29–37.
- 47 Ashizawa, N., Takagi, M., Seto, S. *et al.* (2007) Serum adiponectin and leptin in a patient with Cushing's syndrome before and after adrenalectomy. *Internal Medicine*, **46**, 383–385.
- 48 Berthiaume, M., Sell, H., Lalonde, J. *et al.* (2004) Actions of PPAR γ agonism on adipose tissue remodelling, insulin sensitivity and lipaemia in the absence of glucocorticoids. *American Journal of Physiology. Integrative and Comparative Physiology*, **287**, R1116–R1123.
- 49 Rodriguez-Cuenca, S., Monjo, M., Frontera, M. *et al.* (2007) Sex steroid receptor expression profile in brown adipose tissue. Effects of hormone status. *Cellular Physiology and Biochemistry*, **20**, 877–886.
- 50 Rodriguez, A.M., Monjo, M., Roca, P. *et al.* (2002) Opposite actions of testosterone and progesterone on UCP1 mRNA expression in cultured brown adipocytes. *Cellular and Molecular Life Sciences*, **59**, 1714–1723.
- 51 Rodriguez-Cuenca, S., Monjo, M., Gianotti, M. *et al.* (2007) Expression of mitochondrial biogenesis signalling factors in brown adipocytes is influenced specifically by 17 β oestradiol, testosterone and progesterone. *American Journal of Physiology. Endocrinology and Metabolism*, **292**, E340–E346.
- 52 Monjo, M., Rodriguez, A.M., Palou, A. *et al.* (2003) Direct effects of testosterone, 17 β oestradiol and progesterone on adrenergic regulation in cultured brown adipocytes: a potential mechanism for gender-dependent thermogenesis. *Endocrinology*, **144**, 4923–4930.
- 53 Laudenslager, M.L., Wilkinson, C.W., Carlisle, H.J. *et al.* (1980) Energy balance in ovariectomized rats with and without oestrogen replacement. *American Journal of Physiology*, **238**, R400–R405.
- 54 Pedersen, S.B., Bruun, J.M., Kristensen, K. *et al.* (2001) Regulation of UCP1, UCP2 and UCP3 mRNA expression in brown adipose tissue, white adipose tissue and skeletal muscle in rats by oestrogen. *Biochemical and Biophysical Research Communications*, **288**, 191–197.
- 55 Malo, A. & Puerta, M. (2001) Oestradiol and progesterone change β 3-adrenergic receptor affinity and density in brown adipocytes. *European Journal of Endocrinology*, **145**, 87–91.
- 56 Rice, S.P., Zhang, L., Grennan-Jones, F. *et al.* (2010) Dehydroepiandrosterone (DHEA) treatment in vitro inhibits adipogenesis in human omental but not subcutaneous adipose tissue. *Molecular and Cellular Endocrinology*, **320**, 51–57.
- 57 Ryu, J.W., Kim, M.S., Kim, C.H. *et al.* (2003) DHEA administration increases brown fat uncoupling protein 1 in obese OLETF rats. *Biochemical and Biophysical Research Communications*, **303**, 726–731.
- 58 Bjorntorp, P. (1991) Adipose tissue distribution and function. *International Journal of Obesity*, **15**(Suppl 2), 67–81.
- 59 Medeira, M., Neto, L.V., de Lima, G.A. *et al.* (2010) Effects of GH-IGF-1 excess and gonadal status on BMD and body composition in patients with acromegaly. *Osteoporosis International*, **21**, 2019–2025.
- 60 Meinhardt, V., Nelson, A.E., Hansen, J.L. *et al.* (2010) The effect of growth hormone on body composition and physical performance in recreational athletes: a randomized trial. *Annals of Internal Medicine*, **152**, 568–577.
- 61 Grunfeld, C., Thompson, M., Brown, S.J. *et al.* (2007) Recombinant human growth hormone to treat HIV-associated adipose redistribution syndrome: 12 week induction and 24 week maintenance therapy. *Journal of Acquired Immune Deficiency Syndromes*, **45**, 786–797.
- 62 Li, Y., Knapp, J.R. & Kopchick, J.J. (2002) Enlargement of interscapular brown adipose tissue in growth hormone antagonist transgenic mice and in growth hormone receptor gene-disrupted mice. *Experimental Biology and Medicine*, **228**, 207–215.
- 63 Hioki, C., Yoshida, T., Kogure, A. *et al.* (2004) Effects of growth hormone (GH) on mRNA levels of uncoupling proteins 1, 2 and 3 in brown and white adipose tissues and skeletal muscle in obese mice. *Hormone and Metabolic Research*, **36**, 607–613.
- 64 Lorenzo, M., Valverde, A.M., Teruel, T. *et al.* (1993) IGF-1 is a mitogen involved in differentiation-related gene expression in foetal rat brown adipocytes. *Journal of Cell Biology*, **123**, 1567–1575.
- 65 Valverde, A.M., Mur, C., Brownlee, M. *et al.* (2004) Susceptibility to apoptosis in insulin-like growth factor 1 receptor-deficient brown adipocytes. *Molecular Biology of the Cell*, **15**, 5101–5117.
- 66 Yamashita, H., Kizaki, T., Ookawara, T. *et al.* (1994) Is Insulin-like growth factor 1 involved in brown adipose tissue enlargement? *Life Sciences*, **55**, 141–148.
- 67 Duchamp, C., Burton, K.A., Geleon, A. *et al.* (1997) Transient upregulation of IGF-1 gene expression in brown adipose tissue of cold-exposed rats. *American Journal of Physiology*, **272**, E453–E460.
- 68 Hernandez, A. & Obregon, M.J. (1995) Presence of growth factors-induced type III iodothyronine 5-deiodinase in cultured brown adipocytes. *Endocrinology*, **136**, 4543–4550.

- 69 Vettor, R. & Pagano, C. (2009) The role of the endocannabinoid system in lipogenesis and fatty acid metabolism. *Best Practice & Research. Clinical Endocrinology & Metabolism*, **23**, 51–63.
- 70 Verty, A.M., Allen, A.N. & Oldfield, B.J. (2009) The effects of rimobabant on brown adipose tissue in rat: implications for energy expenditure. *Obesity*, **17**, 254–261.
- 71 Perwitz, N., Wenzel, J., Wagner, I. *et al.* (2010) Cannabinoid type 1 receptor blockade induces transdifferentiation toward a brown fat phenotype in white adipocytes. *Diabetes, Obesity & Metabolism*, **12**, 158–166.
- 72 Rothwell, N.J. & Stock, M.J. (1979) A role for brown adipose tissue in diet-induced thermogenesis. *Nature*, **281**, 31–35.
- 73 Fruhbeck, G., Becerril, S., Sainz, N. *et al.* (2009) BAT: a new target for human obesity? *Trends in Pharmacological Sciences*, **30**, 387–396.
- 74 Hunter, G.R. & Byrne, N.M. (2005) Physical activity and muscle function but not resting energy expenditure impact on weight gain. *Journal of Strength and Conditioning Research*, **19**, 225–230.
- 75 Costford, S., Gowing, A. & Harper, M.E. (2007) Mitochondrial uncoupling as a target in the treatment of obesity. *Current Opinion in Clinical Nutrition and Metabolic Care*, **10**, 671–678.
- 76 Kozak, L.P. & Koza, R.A. (1999) Mitochondrial uncoupling proteins and obesity: molecular and genetic aspects of UCP-1. *International Journal of Obesity*, **23**, S33–S37.
- 77 Zhou, Z., Yon Toh, S., Chen, Z. *et al.* (2003) *Cidea*-deficient mice have lean phenotype and are resistant to obesity. *Nature Genetics*, **35**, 49–56.
- 78 Cederberg, A., Gronning, L.M., Ahren, B. *et al.* (2001) *FOXC2* is a winged helix gene that counteracts obesity, hypertriglyceridaemia and diet-induced insulin resistance. *Cell*, **106**, 563–573.
- 79 Debevec, D. (2007) Receptor interacting protein 140 regulates expression of uncoupling protein 1 in adipocytes through specific peroxisome proliferator activated receptor isoforms and oestrogen-related receptor alpha. *Molecular Endocrinology*, **21**, 1581–1592.
- 80 Timmons, J.A., Wennmalm, K., Larsson, O. *et al.* (2007) Myogenic gene expression signature establishes that brown and white adipocytes originate from distinct cell lineages. *Proceedings of the National Academy of Sciences of the United States of America*, **104**, 4401–4406.
- 81 Charriere, G., Cousin, B., Arnaud, E. *et al.* (2003) Preadipocyte conversion to macrophage – evidence of plasticity. *The Journal of Biological Chemistry*, **278**, 9850–9855.